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Forsmark site investigation

Vegetation inventory in part of the municipality of Östhammar

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July 2003

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Keywords: vegetation inventory, bottom layer, field layer, bush layer, tree layer, fungi

This report concerns a study which was conducted in part for SKB. The conclusions and viewpoints presented in the report are those of the author and do not necessarily coincide with those of the client.

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1 Introduction

A site investigation is an important step in the process of siting a deep repository for spent nuclear fuel. In spring 2001 SKB was interested in conducting thorough investigations in three municipalities; Oskarshamn, Tierp and Östhammar. Since April 2002 Tierp is not longer in the process. Each site investigation is divided into discipline-specific programmes for a number of disciplines. The discipline-specific programme for surface ecosystems aims at an all-round identification and characterization of the surface ecosystems for a comprehensive assessment of the biosphere conditions in the area. The site investigations of surface ecosystems are also supposed to furnish the information on area conditions that enable the site investigations to be carried out in consideration of nature conservation and environmental protection.

One part of the surface ecosystem programme is a general inventory of the area's vegetation types which is carried out in the initial phase of the site investigation. An estimate of the distribution of the area's biotopes as well as the distribution of dominating species is made and presented in GIS format. Existing information on the total quantity (biomass) of the dominant species will later be compiled and calculated for different entities using the vegetation maps. Based on the biomass determination, the annual production of biomass will be calculated enabling estimation of material flows of carbon, water and nutrients. The original description of the vegetation will also be used as a base-line from which long-time monitoring can be performed. The vegetation mapping of the three areas started in the summer of 2001 and continued during 2002.

The activity was performed according to Activity plan, SKB AP PF 400-02-20 (SKB internal controlling document). This report describes the methods used and the results obtained from the inventories in Östhammar. Data from the inventory in Oskarshamn will be published in a separate report. Data from the inventory in Tierp have been incorporated in the database SICADA but no separate report has been written.

2 Material and methods

2.1 The Forsmark area

The Forsmark area (Östhammar community) is situated south-west of the Forsmark nuclear power plant, see Figure 2-1. The area is located in the hemiboreal zone /Ahti et al, 1968/. The forests are dominated by conifers but deciduous trees are rather common, especially in the vicinity of water. The shore displacement is a factor strongly forming the landscape. The lower parts are slowly changing from sea bays to fens as the land uplift proceeds.

The soils are mostly fine grained moraines. They harbour a large share of calcium, which was transported by the land ice from calcareous bedrock further north-east.

The land is mostly covered by forest. However, open acres and grazed pastures dominate.

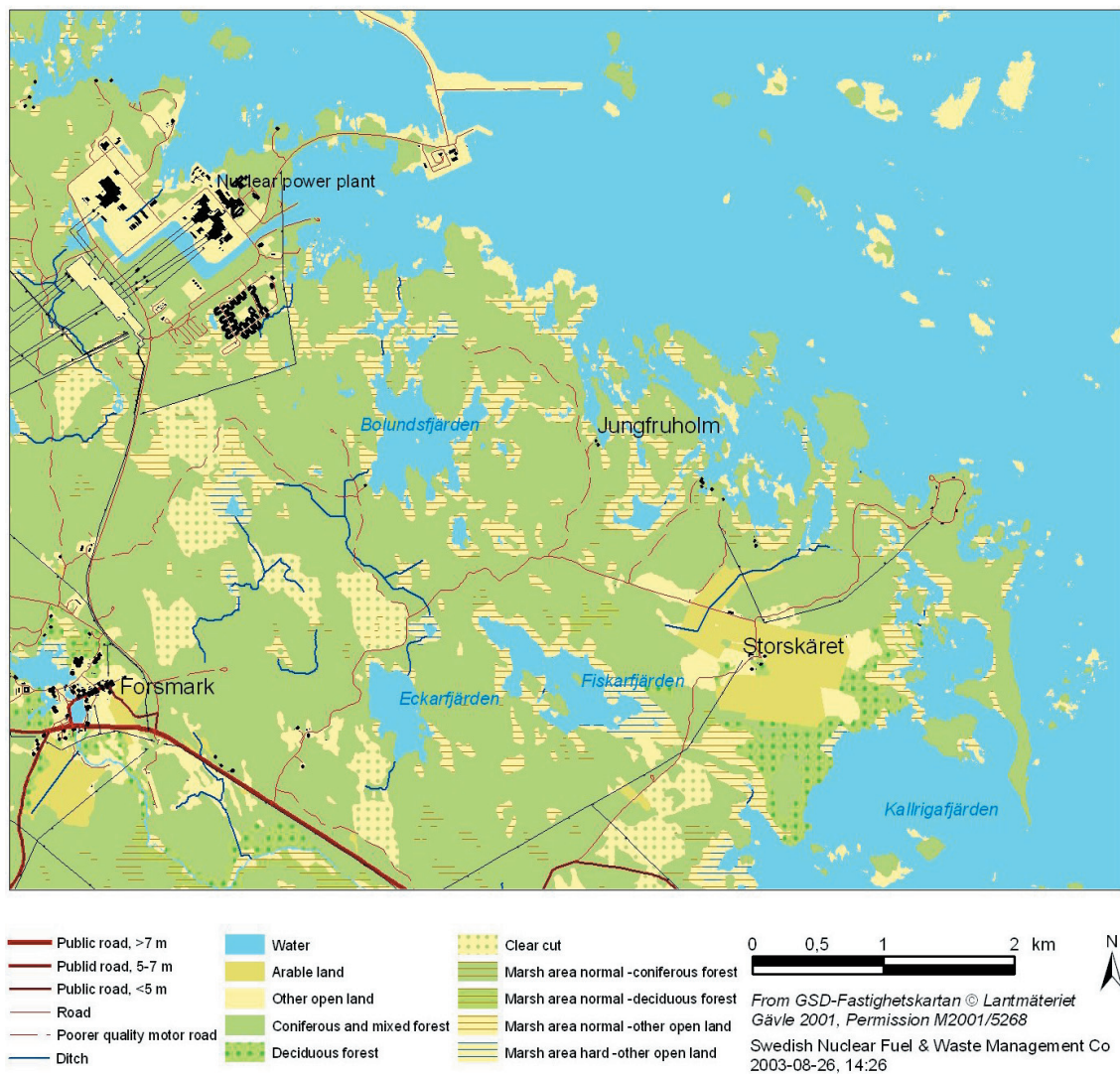


Figure 2-1. The Forsmark area.

2.2 Investigation methods

2.2.1 Identification of vegetation sample areas

The sampling sites were identified using a method with double purposes. The method gives detailed vegetation data and provides monitoring possibilities. The sampling areas are in some cases connected to the places where the initial boreholes for geological investigations were planned to be located. A circle with a radius of 500 m was drawn around each potential drilling site. In 2002 the inventory was expanded. Three new areas, independent of drilling sites, were selected to collect data from other biotopes. Within some (randomly selected) compartments a large square (30x30 m) was located. To these a number of small squares (1x1 m) were connected as described in the section “Location of sampling squares”. The location of all investigated areas at Forsmark can be seen in Figure 2-2. The number of circle areas and large squares identified is shown in Table 2-1.

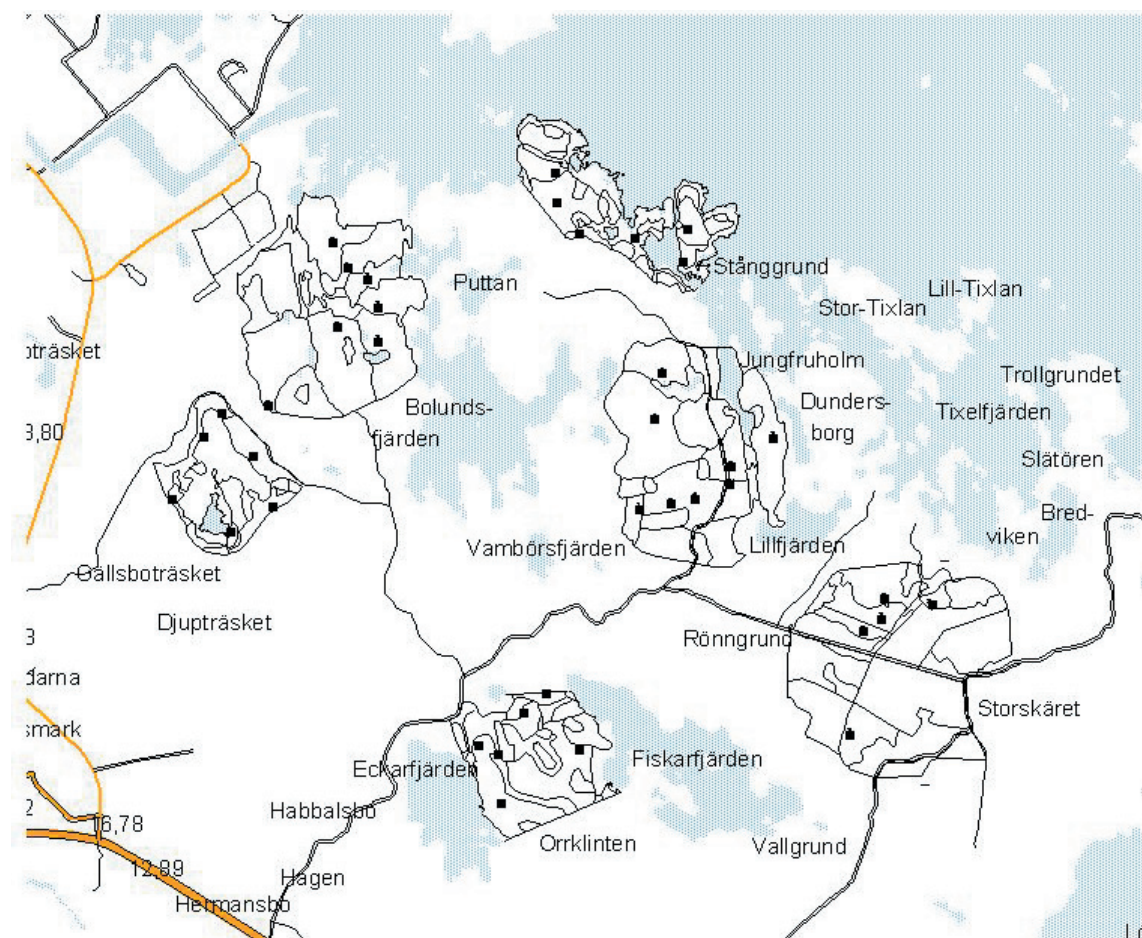


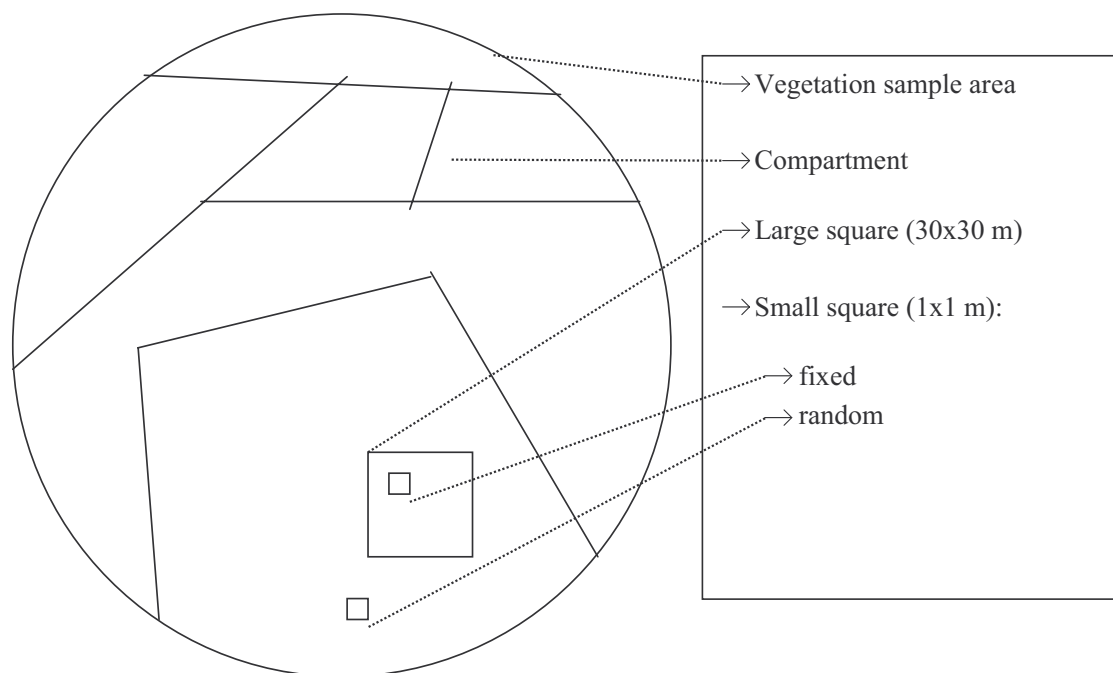
Figure 2-2. The investigated areas at Forsmark. The borders of the compartments are viewed. Large squares are marked with black symbols.

Table 2-1. The different areas investigated during 2001 and 2002 respectively.

Year	Number of circle areas	Number of large squares
2001	3	20
2002	3	18

For each area ortophoto maps were studied. On the map (on screen), heterogeneous parts were divided by borders, so that areas with similar tree characters were delimited in compartments. Thereafter each area was visited. At this visit the borders drawn based on the maps were checked. The field layer characteristics were also studied. If sharp differences in the field layer were discerned, new borders were added. After this visit the borders between the compartments were built on both tree and field layer characteristics. The compartments are thereby describable units in the large circle area (see Figure 2-3). Each compartment was given a description considering dominant species and forest characteristics (site conditions, land use history, conservation values, etc).

In 2002 three new areas were chosen in Forsmark, independent of potential drilling sites. The aim was to gather vegetation data from specific biotopes. Therefore, a wetland, a coastal and a forest area were selected. These vegetation sample areas were given natural borders.



The Vegetation sample area was in 2001 round with a radius of ca 500 metres. In 2002 borders were adjusted to catch a certain vegetation type.

The Compartment is a describable subunit in the circle area with similar tree- and field layer characteristics.

The Large square is 30x30 metre and represents a sample of the compartment.

The Fixed Small squares are samples of the large square. There are nine of them in each large square.

The Random Small squares are randomly placed samples of the compartment. They can be compared to the Fixed Small squares and show whether the Fixed Small squares are representative to the compartment. There are 15 Random Small squares connected to each Large square.

Figure 2-3. The different areas and squares used in the vegetation inventory.

Location of sampling squares

Within some (randomly chosen) compartments a large square was located (30x30 m). Within each large square, nine small squares (1x1 m) were identified. These are hereafter called fixed small squares. Another 15 small squares, located outside the large square but inside the compartment of interest, were also identified. These are hereafter called random small squares. In all, 24 small squares are thus connected to each large square. Exactly the same data were collected in the fixed and random small squares.

The following methods were used when large and small sampling squares were identified. The vegetation sample area were thought of as a system of co-ordinates where the centre point was origo. A number of co-ordinates were randomly selected by computer. These co-ordinates represented the randomly chosen location of the large square (the south-western corner). E.g. the co-ordinates (+210, -152) means that the south-western corner of the square should be positioned 210 metres east and 152 metres south of the centre point. The distance was paced out in field. When arriving at the point, it was considered whether it was very unrepresentative for the compartment it was situated within. If so, the next co-ordinates were checked up.

The fixed small squares were placed within a quadratic spacing (see Figure 2-4). The location of the random small squares was chosen randomly. By setting the south-western corner of the large square as origo, randomly sampled co-ordinates described their location.

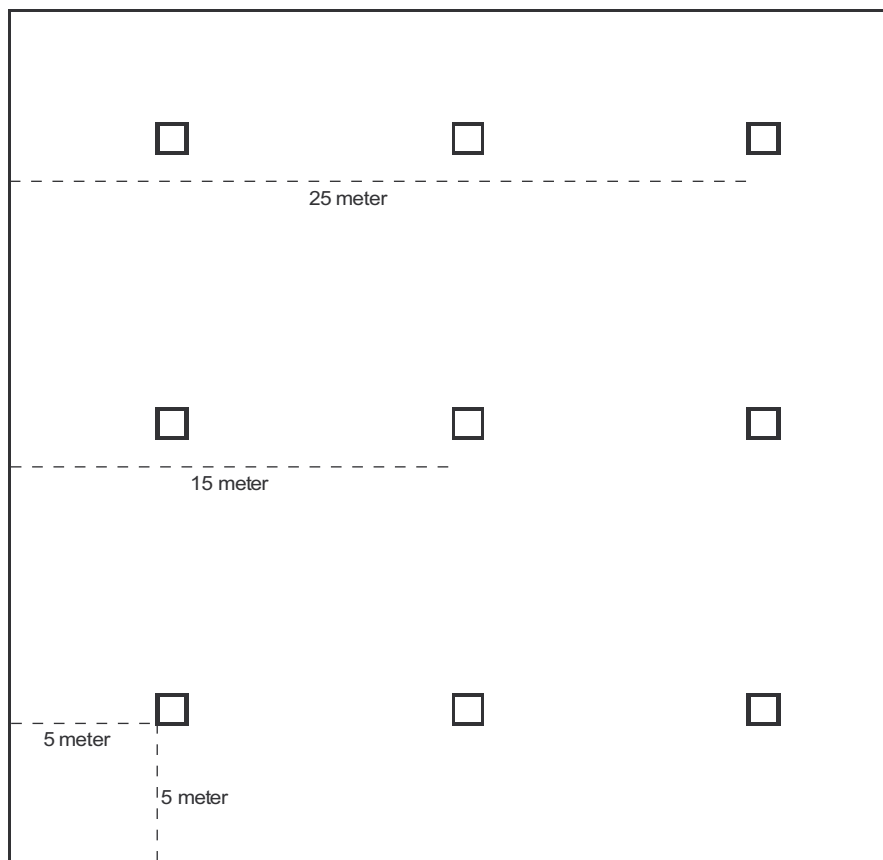


Figure 2-4. The large square (30x30 m) and the fixed small squares (1x1 m). In each corner of the large square a tree stick is placed. In the southwest corner of the fixed small square an aluminium profile is placed, shooting up 10–20 cm. In addition a 15 cm nail is placed below ground level.

Marking of sampling squares

In order to find the sampling squares on the next inventory occasion, both large and small sampling squares have been marked in field. In each corner of the large squares a tree stick has been placed. The small squares are permanently marked, in the south-western corner, underground with a 15 cm nail, and above ground with an aluminium profile emerging 10–20 cm.

2.2.2 Parameters sampled

The sampled parameters are described below. Most parameters have been sampled for both large and small squares, but some were sampled only in small squares, see Table 2-2.

To describe the vegetation in a way that makes it possible to quantify and record changes, **coverage** was considered to be the major unit. The coverage was estimated by studying the vegetation with a vertical projection. If the plants were to be exposed with parallel light from above, the shadowed area would correspond to the coverage. When estimating coverage, all live plant parts above ground are observed. The coverage of bottom, field and bush layer was determined. In the *large squares*, the coverage of the three most dominant bottom layer species was estimated. Furthermore, the coverage of all field layer species covering more than 5% was assessed. However, the coverage of the five most dominant species was always estimated. In the bush layer, the three most common species have been recorded. Presence of other species/genera has been recorded in all layers. In the *small squares* the coverage of the bottom, field, bush and total layer species/genera was estimated. Minimum registration was 1%, which corresponds to 1 dm². This means that all species with a coverage less than 1.5 dm² was given the coverage value 1%.

The coverage of **divergent base** and **substrate** was also estimated. The divergent base includes e.g. large rocks, main haul roads and bare soil from uprooted stumps. The occurrence of such items is supposed to affect the plants long-term access to water and nutrient over long time. The divergent substrate includes elements considered to affect the plants for a short time or to a small degree, e.g. smaller rocks, soft (overgrown) logs and parts of tree roots.

The **diameters** of all trees with a diameter larger than five cm have been measured with a calliper. The **height** of 3–5 trees was measured so that a basal area weighted mean height could be obtained. From the diameter data a basal area was calculated. The basal area and the mean height were input data to achieve a volume. The diameter data were registered per tree species, so that the estimated volume can be distributed on the present tree species.

Furthermore the **litter** was characterised using the following parameters: coverage and thickness of litter layer, litter type and dominating species/group in litter type.

Needle loss was estimated on the tree closest to the south-western corner of each small square. The term “needle loss” represents the lost share of needle mass and this parameter indicates the vitality of the tree. A fully vital tree with no noticeable needle loss is given the value “0%”. A dead tree without needles is given the value “100%”. The method and instruction applied was the same as that used by the National board of forestry. Needle loss was only estimated on pine (*Pinus sylvestris*) and spruce (*Picea abies*) with a breast height diameter exceeding five centimetres. Suppressed or mechanically injured trees were rejected. If no suitable tree was found within five metres of the south-western corner, no needle loss data were registered.

Table 2-2. Data collected at the inventory.

In both large and small squares	Explanation
Divergent base (%)	This class includes e.g. large rocks, main haul roads and bare soil from uprooted stumps. The occurrence of such items is supposed to affect the plants long-term access to water and nutrients.
Bottom layer coverage (%)	The bottom layer consists of bryophytes and lichens.
Field layer coverage (%)	The field layer consists of vascular plants and ferns.
Bush layer coverage (%)	The bush layer consists of ligneous plants with a height between 1–3 metres.
Total layer coverage (%)	The total layer includes bottom, field and bush layer.
Field layer average height (cm)	The average height of the field layer.
Bush layer average height (cm)	The average height of the bush layer.
Litter coverage (%)	The coverage of litter.
In small squares only	
Divergent substrate (%)	This class includes elements considered to affect the plants for a short time or to a minor degree. E.g. smaller rocks, soft (overgrown) logs and parts of tree roots.
Average thickness of litter layer (cm)	
Type of litter	Specifies the dominant kind of litter, e.g. needles, leaves or twigs.
Dominant litter species	The dominant species in litter layer.
Needle loss (%)	The estimated needle loss, indicates the vitality of the tree.
Tree species	Tree species (pine or spruce)
Diameter	Diameter of the tree.
In large squares only	
Tree data	Data collected so that volume per square metre is obtained.

Finally, a brief inventory of **fungi** was made. During five days (early September 2001) two of the sampling sites were searched for redlisted species and species considered as indicators of conservation values. The areas were selected on basis of the observations made during the main inventory work. The field work was carried out by Bo Norell, Foran Sverige AB.

In September 2002 the three new areas were searched for fungi. The remaining area from 2001 was also included.

3 Results

The vegetation inventory of 2001 in Forsmark was carried out by Tommy Abrahamsson and Bo Norell, both working for Foran Sverige AB. It started 2001-07-30 and ended 2001-09-12. The vegetation inventory of 2002 in Forsmark was carried out by Bo Norell. It started in 2002-08-07 and ended 2002-09-15. All data were stored in the database SICADA, the Field Note number is Forsmark 95.

Data from the inventory are presented below. Species lists for the bottom, field, bush, and tree layers can be found in Appendix 1–4. As an example a photo showing one of the fixed small squares is presented in appendix 5 together with a list of the recorded species.

In the following figures the vegetation data have been adjusted to the vegetation maps constructed by SwedPower /Boresjö-Bronge and Wester, 2002/. The large squares have been matched and related to the tree layer code. In Forsmark the most covering tree layer type was “old pine”. Altogether (2001–2002) ten large squares were placed in this tree layer type. The following figures are based on the 15 random small squares associated with each large square.

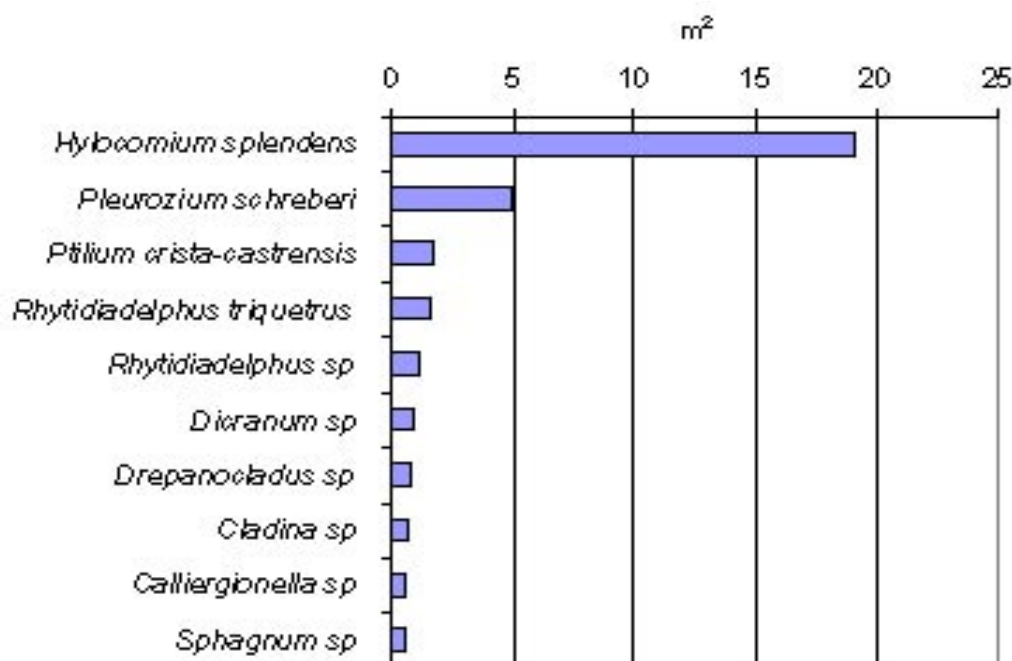
3.1 Bottom layer

In Forsmark altogether (all tree layer types) 36 species/genera have been noted in the bottom layer. The two most dominant species are *Hylocomium splendens* and *Pleurozium schreberi*. They both occurred in one third (301 and 320 of 912) of the total small squares inventoried in Forsmark 2001–2002.

In Table 3-1 the dominant bottom layer species of tree layer type “old pine” are shown. *Hylocomium splendens* is the most covering species. The average cover was 12.7%. It was registered in 71 of the 150 small squares (47%).

Table 3-1. The ten most covering species in the bottom layer, from the tree layer type “old pine”. The sample is based on the random small squares of ten large squares. Sample area is 150 m². *Hylocomium splendens* is the most covering species. It covers 19.12 m² of the sampled 150 m².

Species (Swedish)		Coverage	
		m ²	%
Husmossa	<i>Hylocomium splendens</i>	19.12	12.7
Väggmossa	<i>Pleurozium schreberi</i>	4.95	3.3
Kammossa	<i>Ptilium crista-castrensis</i>	1.74	1.2
Hakmossa	<i>Rhytidiadelphus triquetrus</i>	1.68	1.1
Hakmossor	<i>Rhytidiadelphus sp</i>	1.09	0.7
Kvastmossor	<i>Dicranum sp</i>	0.91	0.6
Krokossor	<i>Drepanocladus sp</i>	0.76	0.5
Renlavar	<i>Cladina sp</i>	0.68	0.5
Spjutmossor	<i>Calliergionella sp</i>	0.65	0.4
Vitmossor	<i>Sphagnum sp</i>	0.60	0.4



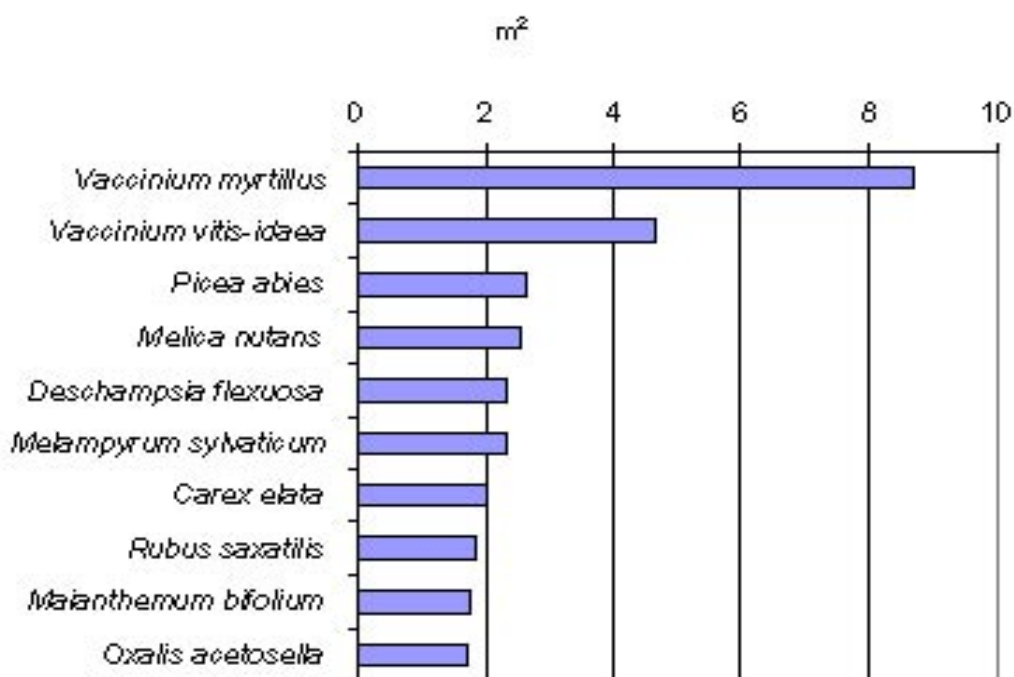
3.2 Field layer

A total of 273 species/genera have been noted in the field layer in the Forsmark area. 17 of these were found in more than ten per cent of the small squares. *Vaccinium myrtillus* was the most covering species. Although *Deschampsia flexuosa* was found in a larger number of squares, it only covered less than half of the area covered by *Vaccinium myrtillus*. 31 of the species have only been noted in one square.

In Table 3-2 the dominant field layer species of tree layer type “old pine” are illustrated. *Vaccinium myrtillus* is the most covering species. It was found in 73 of the 150 small squares (49%). The average cover was 5.8%. The seventh most covering species, *Carex elata* was found in only 19 of the small squares, to be compared with *Oxalis acetosella*, which was identified in 24 small squares.

Table 3-2. The ten most covering species in the field layer from the tree layer type “old pine”. The sample is based on the random small squares of ten large squares. Sample area is 150 m². *Vaccinium myrtillus* is the most covering species. It covers 8.68 m² of the sampled 150 m².

Species (Swedish)		Coverage	
		m ²	%
Blåbär	<i>Vaccinium myrtillus</i>	8.68	5.8
Lingon	<i>Vaccinium vitis-idaea</i>	4.66	3.1
Gran	<i>Picea abies</i>	2.64	1.8
Bergslok	<i>Melica nutans</i>	2.56	1.7
Krustätel	<i>Deschampsia flexuosa</i>	2.35	1.6
Skogskovall	<i>Melampyrum sylvaticum</i>	2.34	1.6
Bunkestarr	<i>Carex elata</i>	2.05	1.4
Stenbär	<i>Rubus saxatilis</i>	1.83	1.2
Ekorrbär	<i>Maianthemum bifolium</i>	1.77	1.2
Harsyra	<i>Oxalis acetosella</i>	1.72	1.1



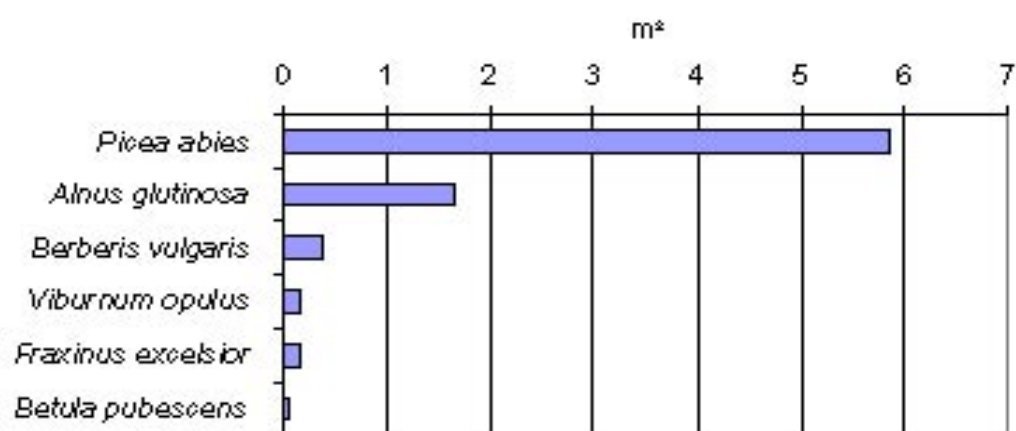
3.3 Bush layer

A total of 19 species/genera have been noted in the bush layer in the Forsmark area. *Picea abies* was the only one that was found in more than ten per cent of the small squares (18%). In Swedish forests the bush layer is normally rather sparsely developed. In Forsmark, bushlayer occurrence (all species) was noted in 16 per cent of the small squares.

In Table 3-3 the six bush layer species of tree layer type “old pine” are illustrated. *Picea abies* is the most covering species. It was found in 20% of the 150 small squares and covered in average 3.9%.

Table 3-3. All registered species in the bush layer from the tree layer type “old pine”. The sample is based on the random small squares of ten large squares. Sample area is 150 m². *Picea abies* is the most covering species. It covers 5.87 m² of the sampled 150 m².

Species (Swedish)		Coverage	
		m ²	%
Gran	<i>Picea abies</i>	5.87	3.9
Klibbal	<i>Alnus glutinosa</i>	1.64	1.1
Berberis	<i>Berberis vulgaris</i>	0.38	0.3
Olvon	<i>Viburnum opulus</i>	0.17	0.1
Ask	<i>Fraxinus excelsior</i>	0.16	0.1
Glasbjörk	<i>Betula pubescens</i>	0.05	0.0



3.4 Tree layer

In Forsmark 14 tree species/genera were noted in the tree layer (Appendix 4). Spruce followed by pine was most often dominating. Tree layer data are calculated from the basal area. With basal area and mean height as input data, a volume per hectare is obtained. This value has then been converted to volume per square metre.

3.5 Fungi

Some interesting species of fungi were found (see Table 3-4), some of them listed as indicator species by the national board of forestry. This means that they are often present at sites with redlisted species. Several of them are considered to indicate sites with high biodiversity and long forest continuity.

Table 3-4. Interesting species of fungi found within the Forsmark area.

<i>Latin name</i>	<i>Swedish name</i>
<i>Cordyceps ophioglossioides</i>	Smal svampklubba
<i>Geastrum sp</i>	Jordstjärna
<i>Cantharellus aurora</i>	Rödgul trumpetsvamp
<i>Phellodon niger</i>	Svart taggsvamp
<i>Hydnellum suavolens</i>	Dofttaggsvamp
<i>Hydnellum ferrugineum</i>	Droptaggsvamp / Skarp droptaggsvamp
<i>Hygrocybe sp</i>	Hagvaxskivling
<i>Sarcodon imbricatus</i>	Fjällig taggsvamp

4 Discussion

4.1 Bottom layer

In the bottom layer *Hylocomium splendens* and *Pleurozium schreberi* are very dominant. Together they constitute 71% of all registered bottom layer coverage in Forsmark.

Whether the field worker is able to do a good description of the bottom layer or not, is strongly affected by the field layer characteristics. In a plot with a well developed field layer it is very difficult to see and extract details of the bottom layer. This is true for the overall bottom layer as well as for single species.

4.2 Field layer

The field layer species exhibit different patterns in nature. The most dominant species (*Vaccinium myrtillus*, *Vaccinium vitis-idea*) are registered in many squares and have high coverage values. Others like *Luzula pilosa* and *Descampsia flexuosa* are noted in many squares but reach only a moderate coverage. *Pteridium aquilinum*, on the other hand, was noted in a few squares, but still reached a high coverage.

Vaccinium myrtillus was the overall dominating species. In average almost 6 per cent of the “old pine” area was covered with this species. The field layer species list has a rather long “tail” with many species of only sparse occurrence.

4.3 Bush layer

When working with squares of 1x1 m, the variation in the bush layer coverage is very high. In “old pine” areas bush layer species only occurred in one square of five.

4.4 Tree layer

The tree layer data is best used as a reference to bottom, field and bush layer data. When basal area is changed, conditions for plants will change too. E.g. if a thinning or a clearcut is performed, plant conditions will be affected. The tree layer data of this survey describes the condition prevailing when vegetation data were collected.

4.5 Fungi

Fungi have been subject to just a brief overview. Anyhow, it is evident that different species of interest are present in these areas. If more intense inventory surveys were carried out, probably additional interesting species would be found. No efforts to quantify fungi biomass have been carried out.

4.6 General

There are several factors influencing the data of this inventory. Some of the main difficulties are discussed below.

- The size difference between the large and the small squares. It is, especially in dense stands, difficult to overview 900 m². Even though segmentation is applied, difficulties are associated with estimating the layer of the large square.
- Whether the large square is representative for the compartment and the variation within the compartment. The data from the small squares can be gathered from an area of several hectares. The mosaic structure of the vegetation may lead to large variations within areas looking homogeneous at a first glance.
- The subjectivity of the field workers. Although the personnel have been calibrated together it is not possible to avoid the influence of subjectivity. Two different field workers most certainly give at least slightly different estimations. This is a part of the method and must be considered when interpreting and using the results.

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Ahti H-A, Jalas J, 1968. Vegetation zones and their sections in northwestern Europe. *Ann. Bot. Fennici*. 5, 169–211.

Boresjö-Bronge L, Wester K, 2002. Vegetation mapping with satellite data of the Forsmark and Tierp regions. SKB report R-02-06.

Inventory manual. Vitality, 1999. National board of forestry.

Bottom layer species/genera noted in small squares in the inventory 2001–2002

<i>Latin names</i>	<i>Swedish names</i>
<i>Aneura pinguis</i>	Fetbålmossa
<i>Antitrichia curtipendula</i>	Fällmossa
<i>Brachythecium sp</i>	Gräsmossor
<i>Calliergion sp</i>	Skedmossor
<i>Calliergionella sp</i>	Spjutmossor
<i>Campylium sp</i>	Spärrmossor
<i>Cladina sp</i>	Renlavar
<i>Cladonia arbuscula</i>	Gulvit renlav
<i>Cladonia rangiferina</i>	Grå renlav
<i>Cladonia sp</i>	Bägarlavar
<i>Cladonia stellaria</i>	Fönsterlav
<i>Climacium dendroides</i>	Palmmossa
<i>Dichelyma sp</i>	Klomossor
<i>Dicranum sp</i>	Kvastmossor
<i>Drepanocladus sp</i>	Krokossor
<i>Helodium blandowii</i>	Kärrkammosa
<i>Herzogiella striatella</i>	Spretmossor
<i>Hylocomium splendens</i>	Husmossa
<i>Hypnum cupressiforme</i>	Cypressfläta (Bergklomossa)
<i>Hypnum sp</i>	Fläta-arter
<i>Hypogymnia sp</i>	Blåslavar
<i>Lophozia sp</i>	Flikmossor
<i>Mnium sp</i>	Stjärnmossor
<i>Peltigera sp</i>	Filtlavar
<i>Plagiochila asplenioides</i>	Praktbräkenmossa
<i>Plagiochila sp</i>	Bräkenmossor
<i>Platismatia sp</i>	Näverlavar
<i>Pleurozium schreberi</i>	Väggmossa
<i>Polytrichum sp</i>	Björnmossor
<i>Pseudotaxiphyllum elegans</i>	Platt skimmermossa
<i>Ptilidium pulcherrinum</i>	Tät franslevermossa
<i>Ptilium crista-castrensis</i>	Kammossa
<i>Rhizocarpon sp</i>	Kartlavar
<i>Rhytidiadelphus sp</i>	Hakmossor
<i>Rhytidiadelphus triquetrus</i>	Kranshakmossa
<i>Scorpidium scorpioides</i>	Korvspionmossa
<i>Sphagnum sp</i>	Vitmossor
<i>Thuidium sp</i>	Tujamossor
<i>Tomentypnum nitens</i>	Gyllenmossa
<i>Warnstorfia exannulata</i>	Kärrkrokmosa

Field layer species/genera noted in small squares in the inventory 2001–2002

<i>Latin names</i>	<i>Swedish names</i>	<i>Latin names</i>	<i>Swedish names</i>
<i>Acer platanoides</i>	Lönn	<i>Carex flacca</i>	Slankstarr
<i>Achillea millefolium</i>	Rölleka	<i>Carex flava</i>	Knagglestarr
<i>Agrimonia eupatoria</i>	Småborre	<i>Carex globularis</i>	Klotstarr
<i>Agrostis canina</i>	Brunven	<i>Carex lasiocarpa</i>	Trådstarr
<i>Agrostis capillaris</i>	Rödven	<i>Carex lepidocarpa</i>	Näbbstarr
<i>Agrostis stolonifera</i>	Krypven	<i>Carex nigra</i>	Hundstarr
<i>Alchemilla sp</i>	Daggkäpa	<i>Carex nigra juncella</i>	Styltstarr
<i>Alnus glutinosa</i>	Klibbal	<i>Carex ovalis</i>	Harstarr
<i>Anemone nemorosa</i>	Vitsippa	<i>Carex pallescens</i>	Blekstarr
<i>Angelica sylvestris</i>	Strätta	<i>Carex panicea</i>	Hirsstarr
<i>Anthoxanthum odoratum</i>	Vårbrodd	<i>Carex pseudocyperus</i>	Slokstarr
<i>Anthriscus sylvestris</i>	Hundkex	<i>Carex pulicaris</i>	Loppstarr
<i>Arrhenatherum elatius</i>	Knylhavre	<i>Carex rostrata</i>	Flaskstarr
<i>Athyrium filix-femina</i>	Majbräken	<i>Carex vesicaria</i>	Blåsstarr
<i>Berberis vulgaris</i>	Berberis	<i>Centarium littorale</i>	Kustarun
<i>Betula pendula</i>	Vårtbjörk	<i>Centaurea jacea</i>	Rödklint
<i>Betula pubescens</i>	Glasbjörk	<i>Cerastium fontanum</i>	Hönsarv
<i>Brachypodium sylvaticum</i>	Lundskafting	<i>Chrysosplenium alternifolium</i>	Gullpudra
<i>Briza media</i>	Darrgräs	<i>Circaea alpina</i>	Dvärghäxört
<i>Calamagrostis arundinacea</i>	Piprör	<i>Cirsium arvense</i>	Åkertistel
<i>Calamagrostis canescens</i>	Grenrör	<i>Cirsium helenioides</i>	Brudborste
<i>Calamagrostis epigejos</i>	Bergrör	<i>Cirsium palustre</i>	Kärrtistel
<i>Calamagrostis purpurea</i>	Brunrör	<i>Cirsium vulgare</i>	Vägtistel
<i>Calamagrostis stricta</i>	Madrör	<i>Coeloglossum viride</i>	Grönkulla
<i>Callitriche stagnalis</i>	Dikeslånke	<i>Convallaria majalis</i>	Liljekonvalj
<i>Caltha palustris</i>	Kabbeleka	<i>Corylus avellana</i>	Hassel
<i>Campanula patula</i>	Ängsklocka	<i>Crepis paludosa</i>	Kärrfibbla
<i>Campanula persicifolia</i>	Stor Blåklocka	<i>Cypripedium calceolus</i>	Guckusko
<i>Cardamine pratensis</i>	Ängsbräsma	<i>Dactylis glomerata</i>	Hundäxing
<i>Carex acuta</i>	Vasstarr	<i>Dactylorhiza fuchsii</i>	Skogsnycklar
<i>Carex appropinquata</i>	Tagelstarr	<i>Dactylorhiza incarnata</i>	Ängsnycklar
<i>Carex canescens</i>	Gråstarr	<i>Danthonia decumbens</i>	Knägräs
<i>Carex capillaris</i>	Hårstarr	<i>Daphne mezereum</i>	Tibast
<i>Carex cespitosa</i>	Tuvstarr	<i>Dapne mezereum</i>	Tibast
<i>Carex cuprina</i>	Blankstarr	<i>Deschampsia cespitosa</i>	Tuvtåtel
<i>Carex diandra</i>	Trindstarr	<i>Deschampsia flexuosa</i>	Kruståtel
<i>Carex digitata</i>	Vispstarr	<i>Dryopteris carthusiana</i>	Skogsbräken
<i>Carex disticha</i>	Plattstarr	<i>Dryopteris cristata</i>	Granbräken
<i>Carex elata</i>	Bunkestarr	<i>Dryopteris filix-mas</i>	Träjon
<i>Carex elongata</i>	Rankstarr	<i>Eleocharis uniglumis</i>	Agnsäv

Latin names	Swedish names	Latin names	Swedish names
<i>Elymus caninus</i>	Lundelm	<i>Hieracium grp silvaticiforma</i>	Skogsfibbla
<i>Elytrigia juncea</i>	Strandkvickrot	<i>Hieracium sp</i>	Hieracium sp
<i>Elytrigia repens</i>	Kvickrot	<i>Hieracium umbellatum</i>	Flockfibbla
<i>Empetrum niger</i>	Kråkbär	<i>Hippophae rhamnoides</i>	Havtorn
<i>Empetrum sp</i>	Kråkbär	<i>Huperzia selago</i>	Lopplummer
<i>Epilobium adenocaulon</i>	Amerikansk dunört	<i>Hypericum maculatum</i>	Fyrkantig Johannesört
<i>Epilobium angustifolium</i>	Mjölkört	<i>Hypericum perforatum</i>	Äkta johannesört
<i>Epilobium palustre</i>	Kärrdunört	<i>Hypochoeris maculata</i>	Slätterfibbla
<i>Epilobium sp</i>	Epilobium sp	<i>Iris pseudacorus</i>	Svärdslilja
<i>Epipactis helleborine</i>	Skogsknipprot	<i>Juncus articulatus</i>	Ryltåg
<i>Epipactis palustris</i>	Kärrknipprot	<i>Juncus balticus</i>	Östersjötåg
<i>Equisetum arvense</i>	Åkerfräken	<i>Juniperus communis</i>	En
<i>Equisetum fluviatile</i>	Sjöfräken	<i>Knautia arvensis</i>	Åkervädd
<i>Equisetum palustre</i>	Kärrfräken	<i>Lactuca sativa</i>	Skogssallat
<i>Equisetum pratense</i>	Ängsfräken	<i>Lathyrus linifolius</i>	Gökärt
<i>Equisetum scirpoides</i>	Trådfräken	<i>Lathyrus pratensis</i>	Gulvial
<i>Equisetum sylvaticum</i>	Skogsfräken	<i>Lathyrus vernus</i>	Vårärt
<i>Eriophorum angustifolium</i>	Ängsull	<i>Ledum palustre</i>	Skvattram
<i>Eupatorium cannabinum</i>	Hampflokel	<i>Lemna minor</i>	Andmat
<i>Euphrasia frigida</i>	Fjällögontröst	<i>Leontodon autumnalis</i>	Höstfibbla
<i>Euphrasia sp</i>	Euphrasia sp	<i>Leucanthemum vulgare</i>	Prästkrage
<i>Festuca arundinacea</i>	Rörsvingel	<i>Linaria vulgaris</i>	Gulmåra
<i>Festuca ovina</i>	Fårsvingel	<i>Linnaea borealis</i>	Linnea
<i>Festuca rubra</i>	Rödsvingel	<i>Listera ovata</i>	Tvåblad
<i>Festuca sp</i>	Festuca sp	<i>Lithospermum officinale</i>	Stellaria sp
<i>Filipendula ulmaria</i>	Älggräs	<i>Lonicera xylosteum</i>	Skogstry
<i>Filipendula vulgaris</i>	Brudbröd	<i>Luzula pallescens</i>	Blekfryle
<i>Fragaria vesca</i>	Smultron	<i>Luzula pilosa</i>	Vårfryle
<i>Frangula alnus</i>	Brakved	<i>Lycopodium annotinum</i>	Revlummer
<i>Fraxinus excelsior</i>	Ask	<i>Lycopus europaeus</i>	Strandklo
<i>Galeopsis bifida</i>	Toppdån	<i>Lysimachia thyrsoiflora</i>	Topplösa
<i>Galium album</i>	Stormåra	<i>Lysimachia vulgaris</i>	Videört
<i>Galium boreale</i>	Vitmåra	<i>Lythrum salicaria</i>	Fackelblomster
<i>Galium palustre</i>	Vattenmåra	<i>Maianthemum bifolium</i>	Ekorrbär
<i>Galium sp</i>	Galium sp	<i>Malus sylvestris</i>	Vildapel
<i>Galium uliginosum</i>	Vattenmåra	<i>Melampyrum nemorosum</i>	Natt och dag
<i>Galium verum</i>	Gulmåra	<i>Melampyrum sylvaticum</i>	Skogskovall
<i>Gentianella uliginosa</i>	Sumpgentiana	<i>Melica nutans</i>	Bergsslok
<i>Geranium robertianum</i>	Stinknäva	<i>Mentha aquatica</i>	Vattenmynta
<i>Geranium sanguineum</i>	Blodnäva	<i>Mentha arvensis</i>	Åkermynta
<i>Geranium sylvaticum</i>	Midsommarblomster	<i>Menyanthes trifoliata</i>	Vattenklöver
<i>Geum rivale</i>	Humbleblomster	<i>Milium effusum</i>	Hässlebrodd
<i>Glaux maritima</i>	Strandkrypa	<i>Molinia caerulea</i>	Blåtåtel
<i>Glechoma hederacea</i>	Jordreva	<i>Monotropa hypopitys</i>	Tallört
<i>Glyceria fluitans</i>	Mannagräs	<i>Myosotis sylvatica</i>	Skogsförgätmigej
<i>Goodyera repens</i>	Knärot	<i>Myrica gale</i>	Pors
<i>Gymnocarpium dryopteris</i>	Ekbräken	<i>Ophioglossum vulgatum</i>	Ormtunga
<i>Hepatica nobilis</i>	Blåsippa	<i>Origanum vulgare</i>	Kungsmynta

Latin names	Swedish names	Latin names	Swedish names
<i>Orthilia secunda</i>	Björkpyrola	<i>Rubus caesius</i>	Blåhallon
<i>Oxalis acetosella</i>	Harsyra	<i>Rubus idaeus</i>	Hallon
<i>Paris quadrifolia</i>	Ormbär	<i>Rubus saxatilis</i>	Stenbär
<i>Parnassia palustris</i>	Slätterblomma	<i>Rubus sp</i>	Rubus sp
<i>Pedicularis palustris</i>	Kärrspira	<i>Rumex acetosa</i>	Ängssyra
<i>Peucedanum palustre</i>	Kärrsilja	<i>Sagittaria natans</i>	Trubbpilblad
<i>Phegopteris connectilis</i>	Hultbräken	<i>Salix aurita</i>	Bindvide
<i>Phleum pratense</i>	Timotej	<i>Salix caprea</i>	Sälg
<i>Phragmites australis</i>	Vass	<i>Salix cinerea</i>	Gråvide
<i>Picea abies</i>	Gran	<i>Salix myrsinifolia</i>	Svartvide
<i>Pimpinella saxifraga</i>	Bockrot	<i>Salix pentandra</i>	Jolster
<i>Pinus sylvestris</i>	Tall	<i>Salix repens</i>	Krypvide
<i>Plantago lanceolata</i>	Svartkämpar	<i>Salix sp</i>	Salix sp
<i>Plantago maritima</i>	Gulkämpar	<i>Sanicula europaea</i>	Sårläka
<i>Platanthera bifolia</i>	Nattviol	<i>Satureja vulgare</i>	Bergmynta
<i>Platanthera chlorantha</i>	Grönvit nattviol	<i>Satureja vulgaris</i>	Bergmynta
<i>Poa nemoralis</i>	Lundgröe	<i>Schoenoplectus lacustris</i>	Säv
<i>Poa palustris</i>	Sengröe	<i>Scrophularia nodosa</i>	Flenört
<i>Poa pratensis</i>	Ängsgröe	<i>Sedum telephium</i>	Kärleksört
<i>Poa trivialis</i>	Kärrgröe	<i>Selaginella selaginoides</i>	Dvärglummer
<i>Polygala vulgaris</i>	Jungfrulin	<i>Sesleria caerulea</i>	Älväxing
<i>Polygonatum multiflorum</i>	Storrams	<i>Silene dioica</i>	Rödblära
<i>Polygonatum odoratum</i>	Getrams	<i>Solanum dulcamara</i>	Besksöta
<i>Polypodium vulgare</i>	Stensöta	<i>Solidago virgaurea</i>	Gullris
<i>Populus tremula</i>	Asp	<i>Sonchus arvensis</i>	Åkermolke
<i>Potentilla anserina</i>	Gåsört	<i>Sorbus aucuparia</i>	Rönn
<i>Potentilla erecta</i>	Blodrot	<i>Sorbus intermedia</i>	Oxel
<i>Potentilla palustris</i>	Kräkklöver	<i>Sparganium natans</i>	Dvärgigelknopp
<i>Potentilla reptans</i>	Revfingerört	<i>Stachys sylvatica</i>	Stinksyska
<i>Primula farinosa</i>	Majviva	<i>Stellaria graminea</i>	Grässtjärnblomma
<i>Primula veris</i>	Gullviva	<i>Stellaria longifolia</i>	Skogsstj.blomma
<i>Prunella vulgaris</i>	Brunört	<i>Stellaria nemorum nemorum</i>	Nordlundarv
<i>Prunus padus</i>	Hägg	<i>Succisa pratensis</i>	Ängsvädd
<i>Pteridium aquilinum</i>	Örnbräken	<i>Taraxacum sp</i>	Maskros
<i>Puccinellia capillaris</i>	Saltgräs	<i>Thalictrum flavum</i>	Ängsruta
<i>Puccinellia distans</i>	Grått saltgäs	<i>Thalictrum simplex</i>	Backruta
<i>Pulmonaria obscura</i>	Lungört	<i>Thalictrum sp</i>	Thalictrum sp
<i>Pyrola chlorantha</i>	Grönpyrola	<i>Thelypteris palustris</i>	Kärrbräken
<i>Pyrola rotundifolia</i>	Vitpyrola	<i>Trientalis europaea</i>	Skogsstjärna
<i>Quercus robur</i>	Ek	<i>Trifolium medium</i>	Skogsklöver
<i>Ranunculus acris</i>	Vanlig Smörblomma	<i>Trifolium pratense</i>	Rödklöver
<i>Ranunculus flammula</i>	Ältranunkel	<i>Trifolium sp</i>	Trifolium sp
<i>Ranunculus lingua</i>	Sjöranunkel	<i>Triglochin maritima</i>	Havssälting
<i>Ranunculus repens</i>	Revsmörblomma	<i>Triglochin palustre</i>	Kärrsälting
<i>Ribes alpinum</i>	Måbär	<i>Tussilago farfara</i>	Tussilago
<i>Roegneria canina</i>	Lundelm	<i>Typha latifolia</i>	Bredkaveldun
<i>Rosa canina</i>	Nyponros	<i>Ulmus glabra</i>	Alm
<i>Rosa sp</i>	Rosa sp	<i>Utricularia intermedia</i>	Dybläddra

Latin names	Swedish names
<i>Utricularia vulgaris</i>	Vattenbläddra
<i>Vaccinium myrtillus</i>	Blåbär
<i>Vaccinium oxycoccos</i>	Tranbär
<i>Vaccinium vitis-idaea</i>	Lingon
<i>Valeriana sambucifolia</i>	Strandvänderot
<i>Veronica chamaedrys</i>	Teveronika
<i>Veronica officinalis</i>	Ärenpris
<i>Veronica scutellata</i>	Dyveronika
<i>Viburnum opulus</i>	Olvon
<i>Vicia cracca</i>	Kräkvicker
<i>Vicia sepium</i>	Häckvicker
<i>Vicia sylvatica</i>	Skogsvicker
<i>Viola canina</i>	Ängsviol
<i>Viola mirabilis</i>	Underviol
<i>Viola palustris</i>	Kärrviol
<i>Viola riviniana</i>	Skogsviol

Bush layer species/genera noted in small squares in the inventory 2001–2002

<i>Latin names</i>	<i>Swedish names</i>
<i>Alnus glutinosa</i>	Klibbal
<i>Berberis vulgaris</i>	Berberis
<i>Betula pendula</i>	Vårtbjörk
<i>Betula pubescens</i>	Glasbjörk
<i>Betula sp</i>	Björk (obest)
<i>Corylus avellana</i>	Hassel
<i>Fraxinus excelsior</i>	Ask
<i>Juniperus communis</i>	En
<i>Myrica gale</i>	Pors
<i>Picea abies</i>	Gran
<i>Pinus sylvestris</i>	Tall
<i>Populus tremula</i>	Asp
<i>Rosa sp</i>	Rosor
<i>Salix caprea</i>	Sälg
<i>Salix cinerea</i>	Gråvide
<i>Salix sp</i>	Salix sp
<i>Sorbus aucuparia</i>	Rönn
<i>Ulmus glabra</i>	Alm
<i>Viburnum opulus</i>	Olvon

Tree species noted and measured in the large squares 2001–2002

<i>Latin names</i>	<i>Swedish names</i>
<i>Alnus glutinosa</i>	Klibbal
<i>Betula pendula</i>	Vårtbjörk
<i>Betula pubescens</i>	Glasbjörk
<i>Betula sp</i>	Björk (obest)
<i>Fraxinus excelsior</i>	Ask
<i>Juniperus communis</i>	En
<i>Picea abies</i>	Gran
<i>Pinus sylvestris</i>	Tall
<i>Populus tremula</i>	Asp
<i>Quercus robur</i>	Ek
<i>Salix caprea</i>	Sälg
<i>Salix pentandra</i>	Jolster
<i>Sorbus aucuparia</i>	Rönn
<i>Ulmus glabra</i>	Alm
<i>Viburnum opulus</i>	Olvon

A photo of a small square with vegetation data

The photo shows the fixed small square AFM000633. It is situated in Forsmark and was investigated 2002. Below the photo the registered bottom, field, and bush layer species are listed. The inventory was carried out in early July and the photo was taken two months later. This might explain why no *Melampyrum sylvaticum* is shown on the photo. The *Melampyrum-species* wilt in August.



Bottom layer Species	Coverage (%)
<i>Hylocomium splendens</i>	50
<i>Polytrichum sp</i>	2

Bottom layer Species	Coverage (%)
<i>Picea abies</i>	3

Field layer Species	Coverage (%)
<i>Vaccinium myrtillus</i>	50
<i>Melampyrum sylvaticum</i>	11
<i>Calamagrostis arundinacea</i>	5
<i>Deschampsia flexuosa</i>	4
<i>Vaccinium vitis-idaea</i>	1